

SERPINE1 as an Independent Prognostic Marker and Therapeutic Target for Nicotine-Related Oral Carcinoma

Xiaopeng Guo*  · Zhen Sun*  · Huarong Chen  · Junjun Ling  · Houyu Zhao  · Aoshuang Chang  · Xianlu Zhuo 

Department of Otorhinolaryngology-Head and Neck Surgery, Affiliated Hospital of Guizhou Medical University, Guiyang, China

Objectives. Nicotine is an ingredient of tobacco, and exposure to nicotine increases the risks of various cancers, including oral cancer. Previous studies have focused on the addictive properties of nicotine, but its carcinogenic mechanism has rarely been studied. We aimed to explore the key genes in the process through which nicotine promotes the occurrence and development of oral cancer via data mining and experimental verification.

Methods. This study involved three parts. First, key genes related to nicotine-related oral cancer were screened through data mining; second, the expression and clinical significance of a key gene in oral cancer tissues were verified by bioinformatics. Finally, the expression and clinical significance of the key gene in oral cancer were histologically investigated, and the effects of its expression on cell proliferation, invasion, and drug resistance were cytologically assessed.

Results. *SERPINE1* was identified as the key gene, which was upregulated in nicotine-treated oral cells and may be an independent prognostic factor for oral cancer. *SERPINE1* was enriched in various pathways, such as the tumor necrosis factor and apelin pathways, and was related to the infiltration of macrophages, CD4+T cells, and CD8+T cells. Overexpression of *SERPINE1* was associated with N staging and may be involved in hypoxia, angiogenesis, and metastasis. Knockdown of *SERPINE1* in oral cancer cells resulted in weakened cell proliferation and invasion ability and increased sensitivity to bleomycin and docetaxel.

Conclusion. This study revealed *SERPINE1* as a key gene for nicotine-related oral cancer, indicating that *SERPINE1* may be a novel prognostic indicator and therapeutic target for oral carcinoma.

Keywords. Nicotine; Mouth Neoplasm; Computational Biology; Neoplasm Metastasis; Prognosis

INTRODUCTION

Oral cancer is a malignant tumor that occurs in the oral cavity and adjacent tissues; it is the most common type of head and neck cancer (HNC), and its incidence is gradually increasing [1]. Despite the combined application of multiple treatment methods, the prognosis is still poor, with a 5-year survival rate of less

than 50% [1]. The main risk factors for oral cancer are smoking, alcohol consumption, betel nut chewing, and human papilloma-virus infection, among which smoking is an established factor [2]. Smoking affects human health in several ways and leads to the development of chronic diseases, such as cardiovascular disease and cancer [3].

Nicotine, the main active and addictive ingredient in tobacco (chemical formula $C_{10}H_{14}N_2$), is an alkaloid found in the *Nicotiana tabacum* plant and is highly toxic [4]. Research has focused on its addictive properties, but rarely on its carcinogenic properties [5]. However, studies have shown that the possible carcinogenic effect of nicotine cannot be ignored. For example, nicotine may promote the progression of various cancers by acting on nicotinic choline receptors [6]. The activation of nicotinic acetylcholine receptors may stimulate cell proliferation and inhibit

• Received October 12, 2022

Revised November 8, 2022

Accepted November 27, 2022

• Corresponding authors: Xianlu Zhuo (zhuoxianlu@gmc.edu.cn) and Aoshuang Chang (changaoshuang@gmc.edu.cn)
Department of Otorhinolaryngology-Head and Neck Surgery, Affiliated Hospital of Guizhou Medical University, Guiyang, Guizhou 550004, China
Tel: +86-851-86773947

*These authors contributed equally to this work.

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apoptosis, thus leading to tumorigenesis [7]. Moreover, nicotine exposure promotes cell metastasis and confers drug resistance in HNC [8]. Nevertheless, the carcinogenic mechanism of nicotine is very complex and still unclear.

Aberrant expression of some genes may mediate nicotine-induced carcinogenesis. For example, nicotine exposure can elevate the expression of $\alpha 5$ nicotinic acetylcholine receptors and survivin in lung cells, which plays an important role in the occurrence and development of lung adenocarcinoma [9]. Nicotine exposure can also induce the abnormal expression of *PRX1* and the genes encoding for its interacting proteins *CFL1* and *PPP2R1A*, thereby promoting the transformation of normal oral cells into cancer cells [10]. However, most of the literature has focused on single genes or pathways, which may lead to biases in our understanding of nicotine's oncogenic mechanisms. Therefore, it is necessary to comprehensively study the molecular mechanisms of nicotine, which may improve our understanding of oral cancer prevention and treatment.

In this study, we aimed to explore dysregulated genes associated with nicotine-related oral cancer and screen key genes by data mining. Then, the expression and function of the key genes were further verified through big data analysis and experiments.

MATERIALS AND METHODS

Screening of the key gene in Nicotine-related oral cancer

Screening of the differentially expressed genes

Datasets regarding nicotine-treated oral cells were retrieved from the Gene Expression Omnibus (GEO) database (<https://ncbi.nlm.nih.gov/geo/>). The differentially expressed genes (DEGs) between the nicotine-treated experimental and control groups were obtained by using the GEO2R [11]. The cut-off criteria of adjusted *P*-value <0.05 and \log_2 |fold-change| >1.5 were used for identification of the DEGs.

Functional enrichment analysis of the DEGs

Gene Ontology (GO) and the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analyses were conducted to annotate the functions of the DEGs. The DAVID database (<https://david.ncifcrf.gov>) was used. A *P*<0.05 was considered statistically significant.

HIGHLIGHTS

- There are many studies on nicotine addiction, but the carcinogenic mechanism of nicotine remains unclear.
- Data mining found that *SERPINE1* may be a key gene for nicotine-related oral cancer.
- *SERPINE1* may be related to the proliferation, invasion, and chemosensitivity of cancer cells.

Screening of the hub genes

To screen for the hub genes/proteins, the DEGs were submitted to STRING (<https://cn.string-db.org>) for calculation. The genes with a composite score of more than 0.4 were chosen. The Cytoscape was used for further calculation and visualization. Genes were sorted by degree and betweenness, respectively, and the top 15 genes were considered the hub genes. A Venn diagram was used to obtain the intersection of the two gene sets. The GEPIA tool (<http://gepia.cancer-pku.cn/>) based on The Cancer Genome Atlas (TCGA) database was used for evaluation.

Validation of the key genes through bioinformatics analysis

Expression levels of the key gene in HNC

The HNC cohort from the TCGA database was used. The associations of the key gene (*SERPINE1*) expression with the confounding factors were assessed. Univariate and multivariate Cox regression analysis was applied to verify the prognostic values of *SERPINE1* and clinicopathological factors in HNC. The prognostic value and possible functions of *SERPINE1* in HNC were also assessed. The GEO database, ProggeneV2 (<http://www.progtools.net/gene/>), and TNMplot (<https://tnmplot.com/analysis/>) were used.

Prediction of intergenic interactions of *SERPINE1*

The NetworkAnalyst tool (<https://networkanalyst.ca>) was used to predict the target genes of *SERPINE1*.

Immune correlation analysis of *SERPINE1*

The TIMER (<http://timer.cistrome.org/>) and CIBERSORT (<https://cibersortx.stanford.edu>) algorithms were used to investigate whether *SERPINE1* expression was related to the tumor immune microenvironment.

Single-cell functional analysis

The CancerSEA database [12] was used to learn the roles of *SERPINE1* in individual HNC cells.

Drug sensitivity prediction

The possible effect of *SERPINE1* expression on the drug sensitivity was predicted by using The Genomics of Drug Sensitivity in Cancer (GDSC) database and the Gene Set Cancer Analysis [13] was used to help visualize the results.

Validations of the key genes by experimental assays

The experiment was approved by the Ethics Committee of Guizhou Medical University. The tissue microarray was commercially purchased, and the consent of the patients had been obtained when collecting samples.

Tissue sample analysis

A tissue microarray of oral cancer patients (HOraC060PG01) was purchased from Shanghai Outdo Biotech Company. The char-

acteristic of the patients was described previously [14]. *SERPINE1* protein expression was measured by immunohistochemistry (IHC) assay. IHC staining was performed according to the manufacturer's instructions. Protein staining intensity was scored from 0 to 3, with 0 indicating negative, 1 weak, 2 moderate, and 3 strong. The percentage of positively stained cells was scored from 0 to 4, where 1 represents (1%–25%), 2 (26%–50%), 3 (51%–75%), and 4 (76%–100%). Samples were scored as the product of percent staining and staining intensity, ranging from 0 to 12. A score of not less than 6 was considered “high” and the rest was considered “low.”

Cell culture

CAL27, SAS, HSC-3 (oral cancer cell lines), and DOK (human dysplastic oral keratinocyte) from American Type Culture Collection were stored in the laboratory. Cells were cultured in Dulbecco's modified Eagle medium containing 10% fetal bovine serum at 37 °C in a humidified atmosphere containing 5% CO₂. DOK was intermittently treated with a low dose of Nicotine (1 μM) for 6 months according to the reference [15]. The treated cells were named DOK/NIC.

Cell line analysis

SERPINE1 was overexpressed in oral cancer tissues. Thus, the loss-of-function strategy was used to explore its roles. The RNA interference technique was utilized. Stable *SERPINE1*-knockdown cell lines were established according to the methods described recently [14]. Two cancer cell lines, CAL27 and SAS, were chosen for further exploration. The cells with stable *SERPINE1*-silence were named sh-*SERPINE1*-CAL27 and sh-*SERPINE1*-SAS, whereas the relevant controls were named sh-NC-CAL27 and sh-NC-SAS, respectively.

The mRNA and protein expressions of *SERPINE1* in these cells were detected by qRT-PCR and Western blot assays (anti-PAI1 rabbit monoclonal antibody; Abcam). The cell proliferation was assayed by CCK-8 and colony formation test. The invasive abilities of the cells were evaluated by transwell invasion assay.

Statistical analysis

For continuous variables, differences between groups were assessed using *t*-tests, analysis of variance, or Wilcoxon rank-sum tests, depending on the specific type of data. If ratio comparisons were involved, the chi-square test was chosen. Overall survival curves were calculated using the Kaplan-Meier method, and differences in survival rates were determined using the log-rank test. Multivariate Cox regression analysis was performed when multiple possible clinical factors were considered. A *P*-value or false discovery rate (FDR) of less than 0.05 was considered statistically significant. The statistical analyses were performed using GraphPad Prism 5 (GraphPad Software Inc.)

RESULTS

Screening for the key gene

Screening of the DEGs

GSE89923 [16], a gene expression profile that met the inclusion criteria, was identified. The dataset was retrieved and downloaded from the GEO database. The dataset contained six samples of normal human oral cells exposed to nicotine and six samples of control cells. They were analyzed on the GPL570 platform of Affymetrix Human Genome U133 Plus 2.0 Array. The DEGs were screened from a comparison of the two groups. Finally, 123 upregulated genes and 109 downregulated genes were identified through screening (Fig. 1A).

Functional annotation of the DEGs

We performed a GO analysis to investigate the possible biological processes of the DEGs. The DEGs were enriched in hundreds of terms. Fig. 1B lists the top 20 terms, such as cell adhesion, epidermal development, neutrophil chemotaxis, epithelial cell differentiation, and neutrophil chemotaxis. KEGG analysis showed the pathway terms. Fig. 1C lists the top 20 terms, including pathways in cancer, interleukin (IL)-17 signaling pathway, tumor necrosis factor (TNF) signaling pathway, transcriptional misregulation in cancer, transforming growth factor (TGF)-β signaling pathway, and nuclear factor (NF)-kappa B signaling pathway.

Selection of *SERPINE1* as a key gene

The interactions of the DEGs were further explored using the STRING and Cytoscape software. In the network, genes were sorted by their degree and betweenness values (Fig. 1D and E, Table 1). The top 15 hub genes were selected separately for Venn analysis (Fig. 1F). The intersection contained eight genes: *EGFR*, *CDH1*, *CXCL8*, *S100A7*, *SERPINE1*, *LCN2*, *EP300*, and *P13*. The prognostic values of these genes in HNC were evaluated using the GEPIA tool. *SERPINE1* was eventually identified as a key gene due to its possible effect on the prognosis of HNC patients and its high expression in HNC tissues (Table 1).

Validation of the roles of the key gene in HNC by bioinformatics methods

Expression of *SERPINE1* in pan-cancer and its prognostic value in HNC cancer

The TIMER database was used to analyze the expression of *SERPINE1* in pan-cancer. As displayed in Fig. 2A, the expression of *SERPINE1* was significantly higher in HNC, BRCA, COAD, ESCA, KIRC, READ, and other tumor tissues that in normal tissues, respectively. Next, survival curves based on the TCGA and GSE68585 data, respectively, showed that the overall survival time of the HNC patients bearing high *SERPINE1* expression was shorter than that of patients harboring low *SERPINE1* expression (Fig. 2B and C).

We evaluated the relationship between *SERPINE1* expression

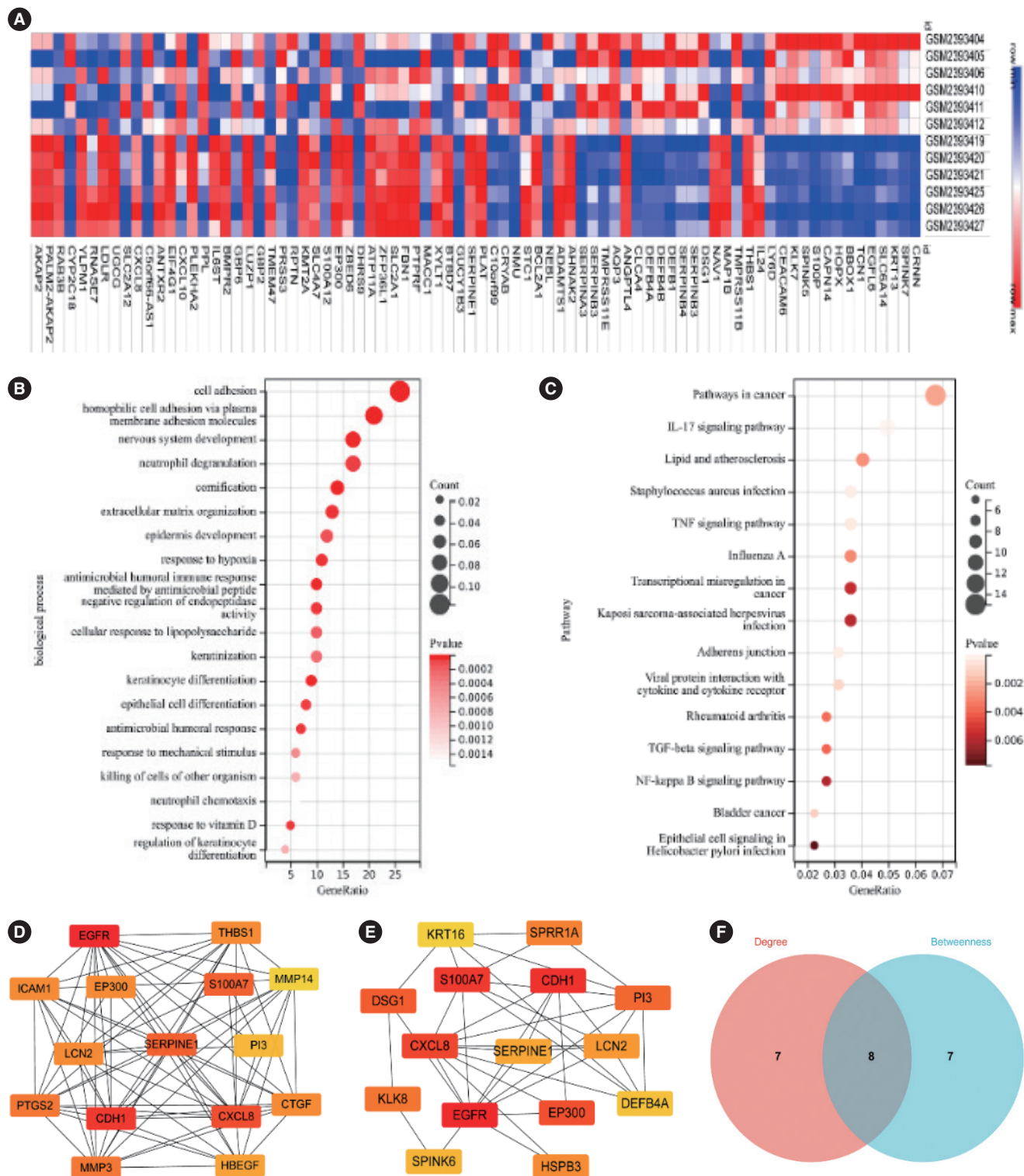


Fig. 1. (A) Heatmap of the differentially expressed genes (DEGs) screened from the GSE89923 dataset. The horizontal axis represents the names of the genes, and the right vertical axis represents the samples. Red represents the upregulated genes and blue represents the down-regulated ones. (B, C) Enrichment analysis of the DEGs by Gene Ontology (GO; B) and Kyoto Encyclopedia of Genes and Genomes (KEGG; C). (D, E) The top 15 genes in a protein-protein interaction network of the DEGs are sorted by degree (D) and betweenness (E) values. (F) Venn analysis of the top 15 hub genes ranked by the degree and betweenness values, respectively. The intersection contained eight genes. IL, interleukin; TNF, tumor necrosis factor; TGF, transforming growth factor; NF, nuclear factor.

Table 2. Associations between *SERPINE1* expression and clinicopathological factors in oral cancer (TCGA)

| Feature | Classification | Number | <i>SERPINE1</i> expression | Z | P-value |
|-----------------------|----------------|--------|----------------------------|-------|---------|
| Age | >65 yr | 129 | 12.618 (6.527–16.937) | 1.145 | 0.252 |
| | ≤65 yr | 213 | 12.764 (8.367–16.545) | | |
| Sex | Female | 106 | 12.947 (6.527–16.973) | 0.983 | 0.325 |
| | Male | 237 | 12.670 (8.367–16.545) | | |
| T-stage | T1-T2 | 129 | 12.521 (6.527–16.973) | 1.475 | 0.140 |
| | T3-T4 | 203 | 12.832 (8.367–16.545) | | |
| N-stage | N0 | 124 | 12.567 (8.367–16.973) | 2.024 | 0.043 |
| | N1-N3 | 163 | 12.908 (8.597–16.545) | | |
| Number of lymph nodes | <1 | 173 | 12.497 (6.527–16.973) | 3.165 | 0.002 |
| | ≥1 | 170 | 12.948 (8.597–16.948) | | |
| M-stage | M0 | 324 | 12.707 (6.527–16.973) | 0.504 | 0.614 |
| | M1 | 2 | 13.160 (12.013–14.307) | | |
| Histologic grade | G1-G2 | 259 | 12.713 (8.367–16.973) | 0.624 | 0.533 |
| | G3-G4 | 75 | 12.752 (6.527–15.938) | | |
| Clinical stage | I-II | 93 | 12.521 (6.527–16.973) | 0.972 | 0.331 |
| | III-IV | 240 | 12.787 (8.367–16.545) | | |

Values are presented as median (range). *SERPINE1* expression values referred to transcripts per million. TCGA, The Cancer Genome Atlas.

and clinicopathological features in HNC. As shown in Table 2, the expression of *SERPINE1* was significantly associated with the N-stage and the number of metastatic lymph nodes ($P<0.05$), indicating that *SERPINE1* expression might have a correlation with lymph node metastasis (LNM). Multivariate Cox analysis of different variables in HNC patients revealed that *SERPINE1* expression level was an independent prognostic factor for HNC (Fig. 2D). A plot from the TNMplot tool showed that high *SERPINE1* expression might have a relationship with enhanced metastatic ability in oral cancer cells (Fig. 2E).

Intergenic interaction prediction for *SERPINE1*

SERPINE1 may closely interact with 57 genes (Fig. 2F). A GO analysis and a KEGG pathway enrichment analysis were also conducted. The top GO terms included metabolic processes, growth, biological regulation, developmental processes, cellular processes, immune system processes, and cellular component tissue biogenesis (Fig. 2G). The KEGG terms mainly involved transcriptional misregulation in cancer, TGF- β signaling pathway, apelin signaling pathway, cell cycle, and thyroid hormone signaling pathway (Fig. 2H).

Analysis of *SERPINE1* gene expression and the tumor microenvironment

Evidence indicates that immune cells play an important role in tumor development, metastasis, and drug resistance [17]. We further explored the relationship between *SERPINE1* and tumor immune cells. Different algorithms were used to calculate the relationship between the *SERPINE1* expression levels and the infiltration of different immune cells.

As shown in Fig. 3A, significant associations were found between *SERPINE1* expression and the infiltration levels of dendritic

cells, macrophages, CD4⁺ T cells, and CD8⁺ T cells ($P<0.05$). The CIBERSORT algorithm showed that *SERPINE1* expression might correlate with the infiltration levels of CD4⁺ T cells, natural killer T cells, and macrophages in HNC samples (Fig. 3B). The above results confirmed that *SERPINE1* might affect the infiltration of immune cells in the cancer microenvironment.

Correlation analysis of *SERPINE1* with the functional status of cancer cells

To further elucidate the possible roles of *SERPINE1* in monocytes in HNC, the single-cell database CancerSEA was selected for assessment [12]. The results showed that *SERPINE1* was positively correlated with metastasis ($r=0.50$, $P<0.05$), angiogenesis ($r=0.35$, $P<0.05$), hypoxia ($r=0.42$, $P<0.05$), and the epithelial-mesenchymal transition (EMT; $r=0.36$, $P<0.05$) (Fig. 3C).

Drug sensitivity prediction

To predict the possible effect of *SERPINE1* expression on the chemosensitivity of cancer cells, data from the GDSC database were evaluated. As shown in Fig. 3D, *SERPINE1* expression was negatively correlated with the sensitivity of cancer cells to several agents, such as 17-AAG, bleomycin, and CHIR-99021. Among these drugs, bleomycin ($r=-0.347$, FDR<0.01) and docetaxel ($r=-0.346$, FDR<0.01) were clinically used for the treatment of oral cancer. Thus, they were chosen for further validation.

Validation of *SERPINE1* expression in oral cancer by experimental assays

Expression of *SERPINE1* mRNA in nicotine-exposed oral cells and oral cancer cells

The mRNA expression of *SERPINE1* was detected in nicotine-exposed oral cells and oral cancer cells. The results showed that

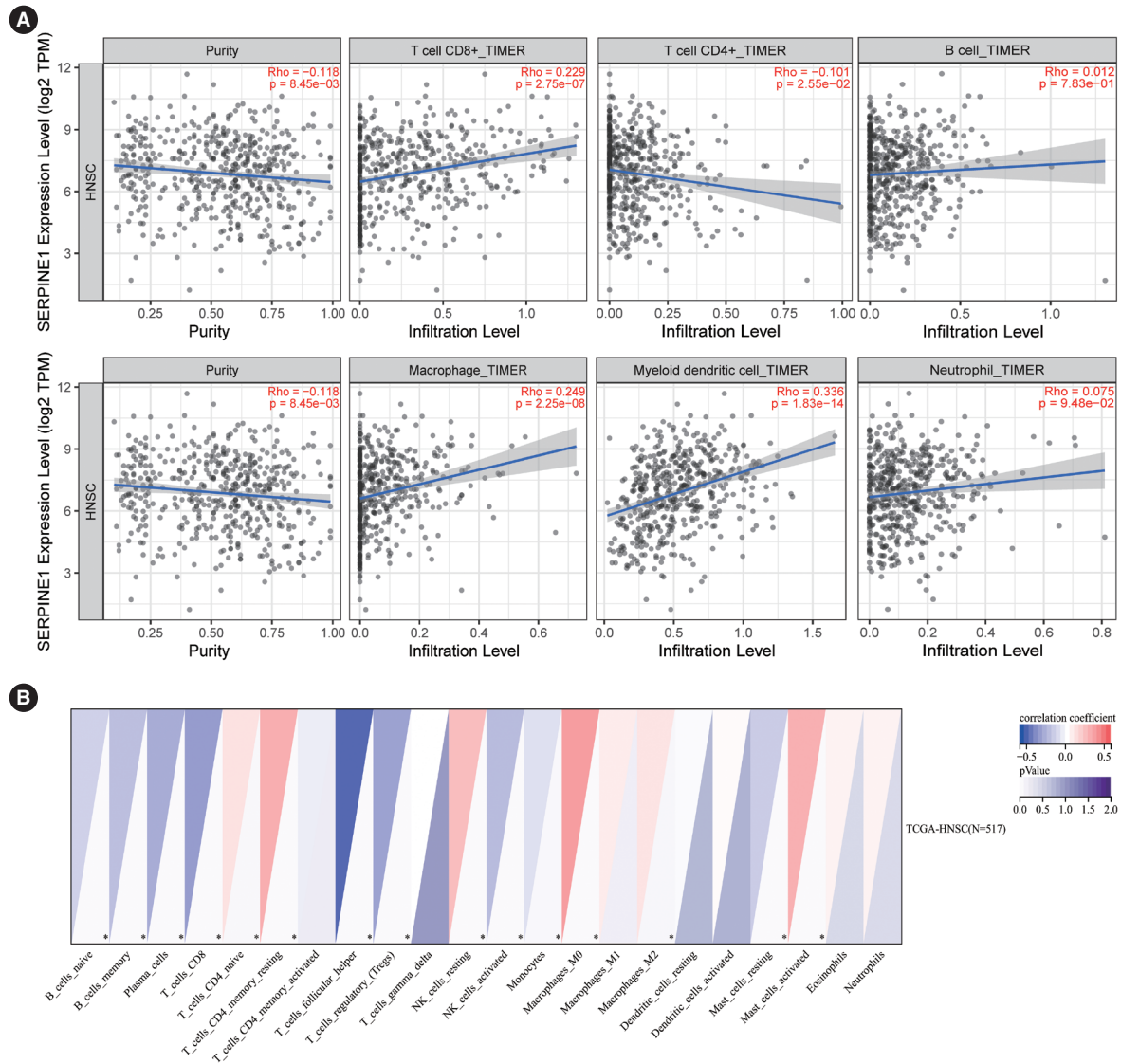


Fig. 3. (A) The association between *SERPINE1* expression and tumor purity, as well as the infiltration levels of several immune cells (TIMER algorithm). (B) Analysis of the relationship between the expression of *SERPINE1* and the infiltration levels of 22 types of immune cells by the Cibersort algorithm. The darker color indicates a higher correlation (* $P < 0.05$). (Continued to the next page)

SERPINE1 mRNA expression was significantly higher in DOK/NIC and oral cancer cell lines than in the control cells (Fig. 4A), suggesting that treatment with nicotine might upregulate *SERPINE1* expression in oral cells and that *SERPINE1* might be an oncogene for oral carcinoma.

Expression of the *SERPINE1* protein in oral cancer tissues

A tissue microarray comprising oral cancer samples was used for detection. Fig. 4B showed that *SERPINE1* protein expression was significantly higher than in oral cancer samples than in controls ($P < 0.05$). *SERPINE1* expression was markedly higher in samples with LNM than in samples without LNM ($P < 0.05$) (Fig. 4C). No associations were identified in comparisons regarding age, sex, and the T-stages (Fig. 4D-G).

Cell proliferation, invasive abilities, and drug sensitivity assessment

Both the mRNA and protein expression levels of *SERPINE1* in the *SERPINE1*-silenced cells were markedly lower than in the control groups ($P < 0.05$) (Fig. 5A and B). The cell proliferation assays showed that the cell viability (Fig. 5C) and colony formation capability (Fig. 5D) were significantly weakened in the *SERPINE1*-knockdown cells (sh-*SERPINE1*-CAL27 and sh-*SERPINE1*-SAS) relative to the controls ($P < 0.05$). Moreover, the invasive ability of the *SERPINE1*-knockdown cells was significantly lower than that of the control cells ($P < 0.05$) (Fig. 5E). The results implied that the overexpression of *SERPINE1* might correlate with enhanced cell proliferation and invasive abilities of oral cancer cells.

To preliminarily verify the effect of *SERPINE1* expression on

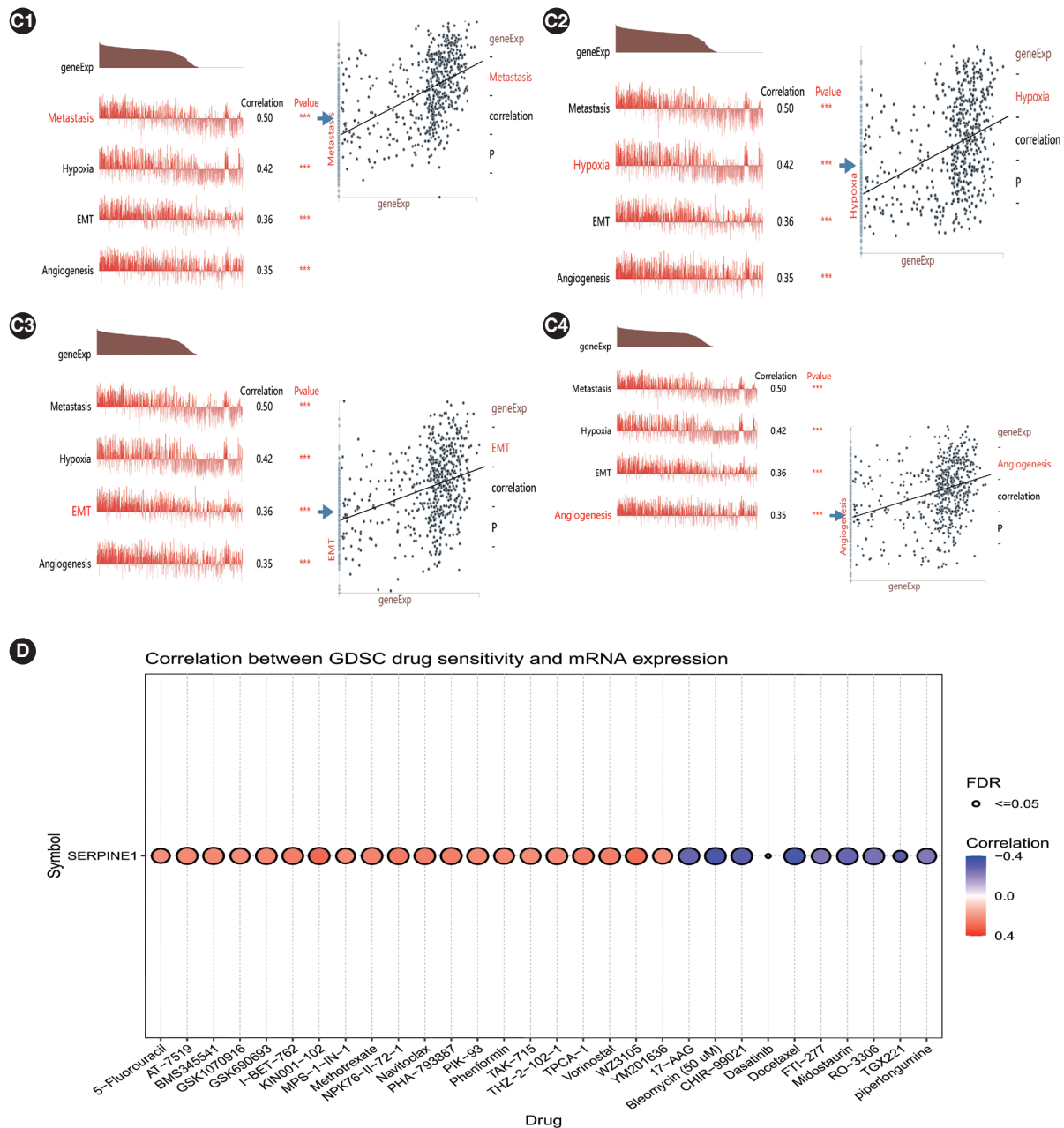


Fig. 3. (Continued) (C) The correlation of *SERPINE1* expression with malignant phenotypes in head and neck cancer tissues. Scatter plots showed positive correlations between *SERPINE1* expression and malignant phenotypes, such as (C1) metastasis, (C2) hypoxia, (C3) the epithelial-mesenchymal transition, and (C4) angiogenesis. (D) The relationship between *SERPINE1* expression and the drug sensitivity of cancer cells. Red represents a positive correlation, while blue stands for a negative correlation. TPM, transcripts per million; TCGA, The Cancer Genome Atlas; HNSC, Head and Neck squamous cell carcinoma; NK, natural killer; EMT, epithelial-mesenchymal transition; GDSC, Genomics of Drug Sensitivity in Cancer; mRNA, messenger RNA; FDR, false discovery rate.

the chemosensitivity of oral cancer cells, cells were treated with 10 μ g/mL bleomycin and 10 nM docetaxel for 48 hours, respectively. The results showed that the cell viability of *SERPINE1*-silenced cells was significantly lower than that of the controls ($P < 0.05$) (Fig. 5F), indicating that the overexpression of *SERPINE1* might confer chemoresistance of oral cancer cells to chemotherapy drugs.

DISCUSSION

In the present study, *SERPINE1* was identified as a key gene that might be involved in nicotine-induced oral carcinogenesis and cancer progression. The overexpression of *SERPINE1* in oral cancer tissues might be correlated with LNM, which predicts poor prognosis. Thus, the downregulation of *SERPINE1* in

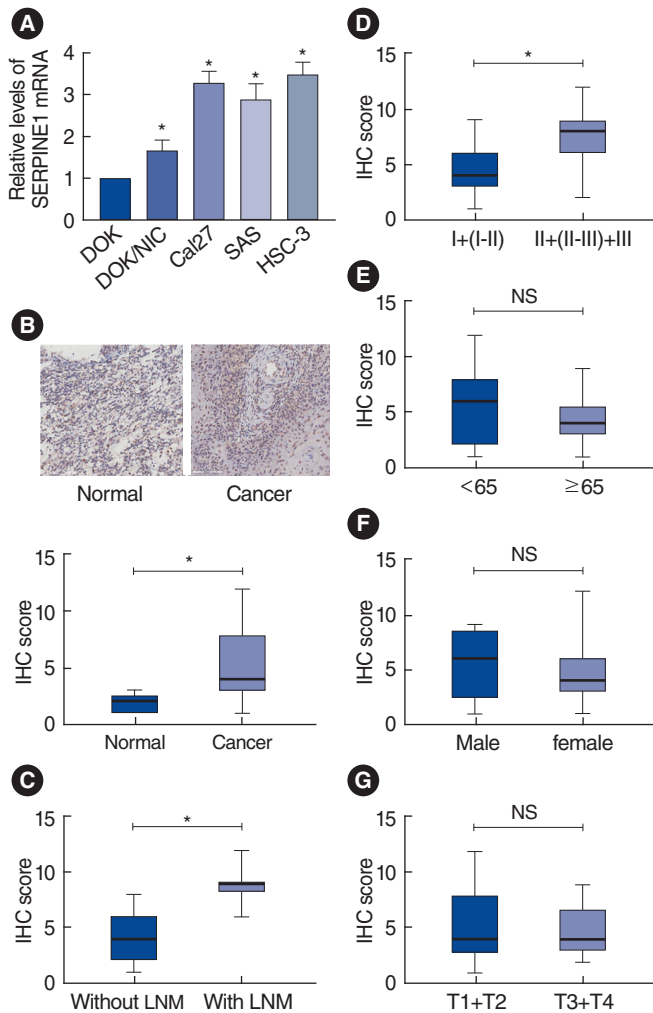


Fig. 4. (A) The messenger RNA (mRNA) expression of *SERPINE1* was higher in nicotine-treated oral cells (DOK/NIC) and oral cancer cell lines (Cal27, SAS, HSC-3) than that in DOK cells, respectively. (B) The immunohistochemistry scores of *SERPINE1* protein expression in oral cancer tissues were markedly higher than in the normal controls. (C) The expression scores for *SERPINE1* protein were higher in cancer samples with lymph node metastasis (LNM) than in those without LNM. (D) The scores of *SERPINE1* expression were higher in the samples with high pathological stages than in those with low stages. No associations were presented concerning age (E), sex (F), and T-stage (G). IHC, immunohistochemistry; NS, not significant ($P > 0.05$). * $P < 0.05$.

oral cancer cells resulted in weakened cell proliferation cell invasion abilities. Moreover, *SERPINE1* silencing resulted in increased cell sensitivity to bleomycin and docetaxel.

SERPINE1, a plasminogen activator inhibitor (PAI-1) protein-coding gene, is located in the long arm of chromosome 7 (7q21.3-q22) and encodes the *SERPINE1* protein, a member of the serine protease inhibitor (serpin) superfamily, which rapidly inhibits fibrinogenesis and mediates a variety of pathological processes such as inflammation and cancer [18]. *SERPINE1* is highly expressed in a variety of tumor tissues [19]. This evidence

is substantially in line with the results of the present study that *SERPINE1* was overexpressed in oral cancer cells and was linked with poor clinical outcomes in patients.

In silico analyses showed that *SERPINE1* expression might be associated with LNM in oral cancer. Single-cell functional analysis indicated that *SERPINE1* expression was positively correlated with hypoxia, the EMT, angiogenesis, and metastasis. The occurrence of LNM in oral cancer patients usually predicts a poor prognosis [20]. Evidence has shown that hypoxia might result in the infiltration of relevant cells, including immune cells, in cancer tissues, which facilitates angiogenesis and thus expedites cancer metastasis [21]. In addition, hypoxia can induce the expression of relevant genes, and then stimulate cells to undergo the EMT through factors such as *FGF1*, thereby promoting angiogenesis [22]. Thus, there seems to be an inseparable relationship among hypoxia, EMT, and angiogenesis, which together promote the invasion and metastasis of cancer cells. Studies showed that a hypoxic environment can induce high expression of *SERPINE1*, which in turn promotes angiogenesis by activation of the EMT process, enhancing the proliferation, invasion, and invasion abilities of cancer cells [23,24]. In the present study, after silencing *SERPINE1* expression in oral cancer cells, we observed significantly weakened cell proliferation and invasion ability. This result suggests that *SERPINE1* expression enhances the malignant properties of oral cancer cells, while targeting *SERPINE1* may significantly reverse the malignant phenotype of the cancer cells.

It is worth noting that *SERPINE1* expression was predicted to have an association with the sensitivity of cancer cells to a series of drugs. Among the significantly related drugs, bleomycin and docetaxel have been applied in the clinical treatment of HNC, and therefore they were selected for further verification. Bleomycin, which has been used in the treatment of HNC, can exert cytotoxic effects in cancer cells by inducing reactive oxygen species production; however, the aberrant expression of some proteins, such as coenzyme Q10, can increase the resistance of oral cancer cells to bleomycin [25]. Docetaxel has been used in first-line chemotherapy for HNC, as well as in the treatment of recurrent and metastatic HNC. Likewise, the emergence of resistance to docetaxel presents a challenge for the treatment of oral cancer [26]. The results of the present study showed that after silencing *SERPINE1* in oral cancer cells, the sensitivity of the cells to both bleomycin and docetaxel significantly increased, suggesting that high expression of *SERPINE1* may reduce the sensitivity of cancer cells to chemotherapy drugs.

To explore the carcinogenic mechanism of nicotine, a functional enrichment analysis of the DEGs related to nicotine-associated carcinogenesis was performed. These genes are enriched in many cancer-related signaling pathways, such as the IL-17 signaling pathway, TNF signaling pathway, and TGF- β signaling pathway. These pathways may play important roles in tumorigenesis and development. For instance, IL-17 promotes tumorigenesis by

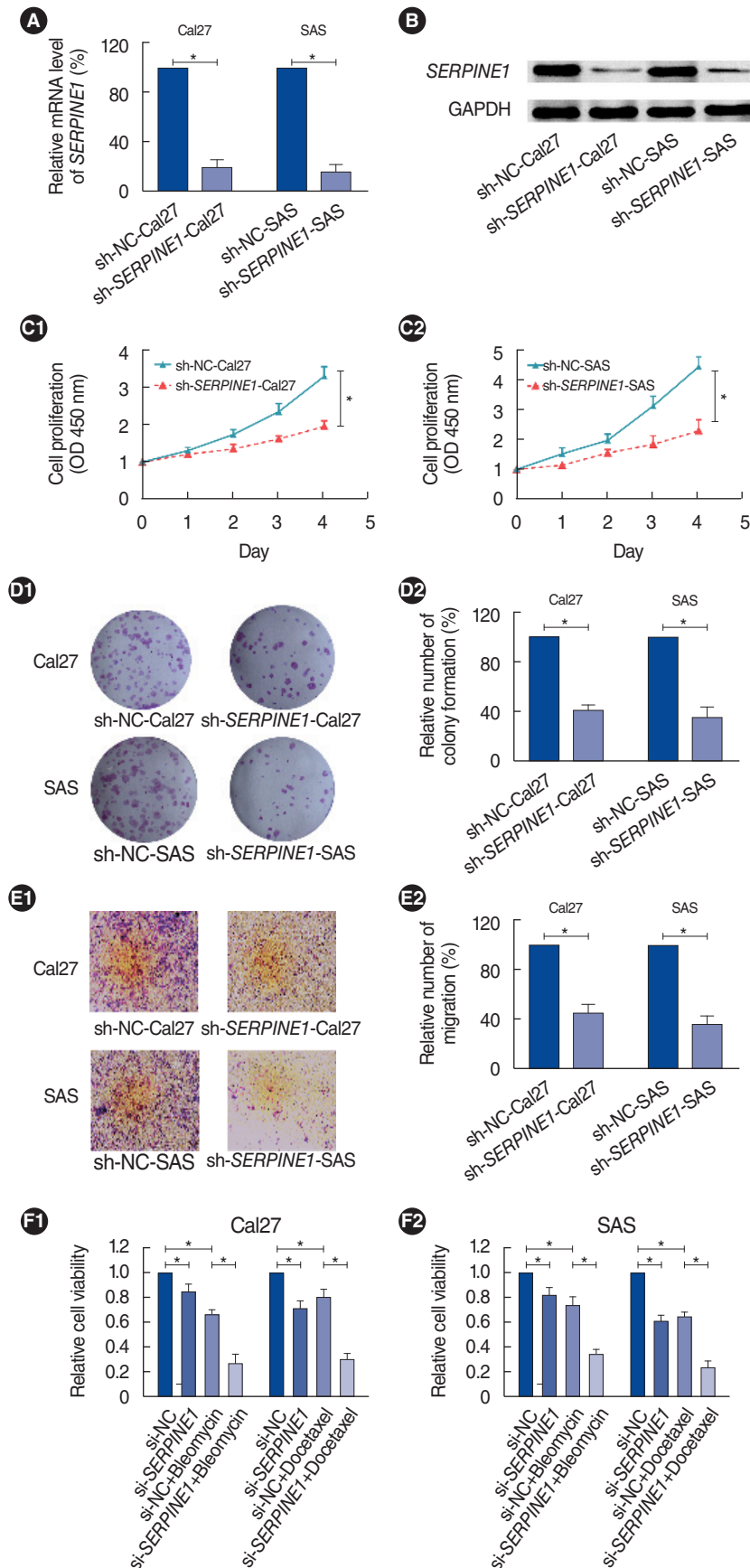


Fig. 5. (A) The mRNA expression of *SERPINE1* was significantly downregulated in the *SERPINE1*-silenced oral cancer cells (sh-*SERPINE1*-Cal27 or sh-*SERPINE1*-SAS) compared with that of the control cells (sh-NC-Cal27 and sh-NC-SAS). (B) The trend of *SERPINE1* protein expression was in line with that of mRNA expression. (C) The cell proliferation abilities of the *SERPINE1*-silenced cancer cells were significantly lower than those of the control cells. (D) The number of colonies formed in the *SERPINE1*-silenced cells was significantly lower than that in the control cells. (E) The invasive abilities in the *SERPINE1*-silenced cells were significantly inhibited compared with those in the control cells. (F) The administration of Bleomycin or docetaxel resulted in a significant decrease in cell viability in *SERPINE1*-silenced cells compared with the control cells. mRNA, messenger RNA; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; NC, negative control; OD, optical density. * $P < 0.05$.

regulating beclin-1 ubiquitination [27]. The TNF signaling pathway mediates the mesenchymal transition of glioblastoma and plays a key role in its progression [28]. The data showed that several signaling pathways with different functions might be involved in the occurrence and development of nicotine-related oral cancer. The genes closely related to *SERPINE1* were also functionally annotated and found to be enriched in multiple signaling pathways. These pathways were scattered across various aspects of cellular functions, and they were not observed to be concentrated in any area. The data suggest that *SERPINE1* may exert biological activities in tumorigenesis and development through multiple different signaling pathways. However, its specific mechanism needs further experimental verification.

We noted that *SERPINE1* expression in oral cancer may be associated with the infiltration of immune cells, including dendritic cells, macrophages, CD4+T cells, and CD8+T cells. The immune microenvironment was involved in the occurrence and progression of cancer. In general, CD8+ T-cell infiltration indicates a better prognosis in several tumors [29], while overexpression of *SERPINE1* predicts a poor prognosis. Nevertheless, a positive correlation between CD8+ T and *SERPINE1* expression was observed. The discrepancy might be because *SERPINE1* promotes the infiltration of CD8+ T cells as only part of its effect, and the influence of this partial effect on the overall prognosis was not sufficient to counteract its other effects (such as the infiltration of tumor-associated macrophages) that may be detrimental to the prognosis. Therefore, the factors influencing tumor prognosis should be comprehensively analyzed. These results suggest that there may be a correlation between *SERPINE1* expression and the infiltration of various immune cells. However, the mechanism by which *SERPINE1* affects immune cell infiltration is unclear and needs to be further clarified in future experimental studies.

Several limitations might exist in the present study. First, the experimental validation only involved *in vitro* assays. The roles of the genes were not verified *in vivo*. Second, the study focused on the effect of nicotine on the oral cavity. Future studies considering the larynx and hypopharynx may enhance our understanding of the carcinogenic effects and mechanisms of nicotine. Third, whether other carcinogens in tobacco such as benzopyrene and nitric oxide can also cause alterations in *SERPINE1* expression is uncertain and was not addressed in this study. Future studies need to explore this issue because it would help to deepen our understanding of the mechanism of nicotine carcinogenesis. Fourth, although this study explored the correlation between nicotine exposure and *SERPINE1* expression through a bioinformatics analysis and cytological verification, the causal relationship between them remains insufficiently understood, and the mechanism through which nicotine regulates *SERPINE1* expression is still unclear. These problems should be taken into account in future experiments.

Despite its limitations, the present study has revealed that

SERPINE1 might be a key gene that plays a crucial role in the genesis and development of nicotine-related oral cancer, raising the possibility that *SERPINE1* could serve as a prognostic factor and a therapeutic target for oral carcinoma. Future experiments are needed to explore the molecular mechanisms in-depth.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

This study was partially supported by the Guizhou Science and Technology Project (ZK2022-044), and the Cultivation project of Affiliated Hospital of Guizhou Medical University (I-2020-10 and gyfysky-2021-60).

ORCID

| | |
|----------------|-------------------------------------------------------------------------------------------|
| Xiaopeng Guo | https://orcid.org/0000-0002-3662-2544 |
| Zhen Sun | https://orcid.org/0000-0003-2122-7960 |
| Huarong Chen | https://orcid.org/0000-0002-8598-3899 |
| Junjun Ling | https://orcid.org/0000-0002-2283-5047 |
| Houyu Zhao | https://orcid.org/0000-0003-2861-0997 |
| Aoshuang Chang | https://orcid.org/0000-0002-1986-2909 |
| Xianlu Zhuo | https://orcid.org/0000-0001-9580-8456 |

AUTHOR CONTRIBUTIONS

Conceptualization: AC, XZ. Data curation: XG, ZS, HC. Formal analysis: XG, JL. Methodology: JL, HZ. Visualization: HZ, XZ. Writing—original draft: JL, XZ, AC. Writing—review & editing: JL, XZ.

REFERENCES

1. Rivera C. Essentials of oral cancer. *Int J Clin Exp Pathol*. 2015 Sep; 8(9):11884-94.
2. Auperin A. Epidemiology of head and neck cancers: an update. *Curr Opin Oncol*. 2020 May;32(3):178-86.
3. Onor IO, Stirling DL, Williams SR, Bediako D, Borghol A, Harris MB, et al. Clinical effects of cigarette smoking: epidemiologic impact and review of pharmacotherapy options. *Int J Environ Res Public Health*. 2017 Sep;14(10):1147.
4. Alkhatib R, Alkhatib B, Abdo N. Impact of exogenous nicotine on the morphological, physio-biochemical, and anatomical characteristics in *Capsicum annum*. *Int J Phytoremediation*. 2022;24(6): 666-74.

5. Murphy SE. Biochemistry of nicotine metabolism and its relevance to lung cancer. *J Biol Chem*. 2021 Jan-Jun;296:100722.
6. Dang N, Meng X, Song H. Nicotinic acetylcholine receptors and cancer. *Biomed Rep*. 2016 May;4(5):515-8.
7. Chen J, Cheuk IW, Shin VY, Kwong A. Acetylcholine receptors: Key players in cancer development. *Surg Oncol*. 2019 Dec;31:46-53.
8. Shimizu R, Ibaragi S, Eguchi T, Kuwajima D, Kodama S, Nishioka T, et al. Nicotine promotes lymph node metastasis and cetuximab resistance in head and neck squamous cell carcinoma. *Int J Oncol*. 2019 Jan;54(1):283-94.
9. Zhang Y, Sun Y, Jia Y, Zhang Q, Zhu P, Ma X. $\alpha 5$ -nAChR and survivin: two potential biological targets in lung adenocarcinoma. *J Cell Physiol*. 2021 Mar;236(3):1787-97.
10. Qi M, Li L, Tang X, Lu Y, Wang M, Yang J, et al. Nicotine promotes the development of oral leukoplakia via regulating peroxiredoxin 1 and its binding proteins. *Braz J Med Biol Res*. 2021 May;54(9):e10931.
11. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, et al. NCBI GEO: archive for functional genomics data sets: update. *Nucleic Acids Res*. 2013 Jan;41(Database issue):D991-5.
12. Yuan H, Yan M, Zhang G, Liu W, Deng C, Liao G, et al. CancerSEA: a cancer single-cell state atlas. *Nucleic Acids Res*. 2019 Jan;47(D1):D900-8.
13. Li X, Yang A, Wen P, Yuan Y, Xiao Z, Shi H, et al. Nuclear receptor subfamily 3 group c member 2 (NR3C2) is downregulated due to hypermethylation and plays a tumor-suppressive role in colon cancer. *Mol Cell Biochem*. 2022 Nov;477(11):2669-79.
14. Feng S, Yuan W, Sun Z, Guo X, Ling J, Chang A, et al. SPP1 as a key gene in the lymph node metastasis and a potential predictor of poor prognosis in head and neck carcinoma. *J Oral Pathol Med*. 2022 Aug;51(7):620-9.
15. Chang YW, Singh KP. Nicotine-induced oxidative stress contributes to EMT and stemness during neoplastic transformation through epigenetic modifications in human kidney epithelial cells. *Toxicol Appl Pharmacol*. 2019 Jul;374:65-76.
16. Woo S, Gao H, Henderson D, Zacharias W, Liu G, Tran QT, et al. AKR1C1 as a biomarker for differentiating the biological effects of combustible from non-combustible tobacco products. *Genes (Basel)*. 2017 May;8(5):132.
17. Zhang T, Yu H, Dai X, Zhang X. CMTM6 and CMTM4 as two novel regulators of PD-L1 modulate the tumor microenvironment. *Front Immunol*. 2022 Jul;13:971428.
18. Chen TY, Zhou M, Lin MQ, Liang ST, Yan Y, Wang SM, et al. Research progress on the SERPINE1 protein and chronic inflammatory diseases of the upper respiratory tract: a literature review. *Int Arch Allergy Immunol*. 2021;182(11):1097-102.
19. Hu B, Chen Z, Wang X, Chen F, Song Z, Cao C. MicroRNA-148a-3p directly targets SERPINE1 to suppress EMT-Mediated colon adenocarcinoma progression. *Cancer Manag Res*. 2021 Aug;13:6349-62.
20. Dai Y, Wang Z, Yan E, Li J, Ge H, Xiao N, et al. Development of a novel signature derived from single cell RNA-sequencing for preoperative prediction of lymph node metastasis in head and neck squamous cell carcinoma. *Head Neck*. 2022 Oct;44(10):2171-80.
21. Sugiura K, Nakajima S, Kato I, Okubo-Sato M, Nakazawa Y, Mitsudo K, et al. Hypoxia and CD11b+ cell influx are strongly associated with lymph node metastasis of oral cancer. *Anticancer Res*. 2020 Dec;40(12):6845-52.
22. Li JP, Liu YJ, Zeng SH, Gao HJ, Chen YG, Zou X. Identification of COX4I2 as a hypoxia-associated gene acting through FGF1 to promote EMT and angiogenesis in CRC. *Cell Mol Biol Lett*. 2022 Sep;27(1):76.
23. Teng B, Xie C, Zhao Y, Zeng Q, Zhan F, Feng Y, et al. Identification of MEDAG and SERPINE1 related to hypoxia in abdominal aortic aneurysm based on weighted gene coexpression network analysis. *Front Physiol*. 2022 Jul;13:926508.
24. Yang JD, Ma L, Zhu Z. SERPINE1 as a cancer-promoting gene in gastric adenocarcinoma: facilitates tumour cell proliferation, migration, and invasion by regulating EMT. *J Chemother*. 2019 Nov-Dec;31(7-8):408-18.
25. Yen HC, Li SH, Majima HJ, Huang YH, Chen CP, Liu CC, et al. Up-regulation of antioxidant enzymes and coenzyme Q(10) in a human oral cancer cell line with acquired bleomycin resistance. *Free Radic Res*. 2011 Jun;45(6):707-16.
26. Cui J, Wang H, Zhang X, Sun X, Zhang J, Ma J. Exosomal miR-200c suppresses chemoresistance of docetaxel in tongue squamous cell carcinoma by suppressing TUBB3 and PPP2R1B. *Aging (Albany NY)*. 2020 Apr;12(8):6756-73.
27. Bie Q, Song H, Chen X, Yang X, Shi S, Zhang L, et al. IL-17B/IL-17RB signaling cascade contributes to self-renewal and tumorigenesis of cancer stem cells by regulating Beclin-1 ubiquitination. *Oncogene*. 2021 Mar;40(12):2200-16.
28. Yan T, Tan Y, Deng G, Sun Z, Liu B, Wang Y, et al. TGF- β induces GBM mesenchymal transition through upregulation of CLDN4 and nuclear translocation to activate TNF- α /NF- κ B signal pathway. *Cell Death Dis*. 2022 Apr;13(4):339.
29. Maimela NR, Liu S, Zhang Y. Fates of CD8+ T cells in tumor microenvironment. *Comput Struct Biotechnol J*. 2018 Nov;17:1-13.